



Endosymbiosis morphological reorganization during metamorphosis diverges in weevils

Justin Maire , Bessem Chouaia , Anna Zaidman-Rémy & Abdelaziz Heddi

To cite this article: Justin Maire , Bessem Chouaia , Anna Zaidman-Rémy & Abdelaziz Heddi (2020) Endosymbiosis morphological reorganization during metamorphosis diverges in weevils, Communicative & Integrative Biology, 13:1, 184-188, DOI: [10.1080/19420889.2020.1840707](https://doi.org/10.1080/19420889.2020.1840707)

To link to this article: <https://doi.org/10.1080/19420889.2020.1840707>



© 2020 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.



Published online: 02 Nov 2020.



Submit your article to this journal [↗](#)



Article views: 217



View related articles [↗](#)



View Crossmark data [↗](#)

Endosymbiosis morphological reorganization during metamorphosis diverges in weevils

Justin Maire^{a,b}, Bessem Chouaia^{c,d}, Anna Zaidman-Rémy^a, and Abdelaziz Heddi^a

^aUniv Lyon, INSA-Lyon, INRA, BF2i, UMR0203, Villeurbanne, France; ^bCurrent address: School of Biosciences, The University of Melbourne, Melbourne, VIC, Australia; ^cDipartimento di Scienze Agrarie e Ambientali (Disaa), Università Degli Studi di Milano, Milan, Italy; ^dCurrent address: Dipartimento di Scienze Molecolari e Nanosistemi (DSMN), Università ca' Foscari di Venezia, Venice, Italy

ABSTRACT

Virtually all animals associate with beneficial symbiotic bacteria. Whether and how these associations are modulated across a host's lifecycle is an important question in disentangling animal-bacteria interactions. We recently reported a case of complete morphological reorganization of symbiosis during metamorphosis of the cereal weevil, *Sitophilus oryzae*. In this model, the bacteriome, a specialized organ that houses the intracellular bacterium *Sodalis pierantonius*, undergoes a two-phase remodeling program synchronously driven by host and endosymbiont, resulting in a localization shift and the formation of multiple new bacteriomes. Here, we provide comparative data in a closely-related coleopteran, the red palm weevil *Rhynchophorus ferrugineus*, which is associated with the ancestral endosymbiont *Nardonella*. Using cell imaging experiments, we show that the red pal weevil bacteriome remains unchanged during metamorphosis, hence contrasting with what we reported in the cereal weevil *S. oryzae*. These findings highlight the complexity and divergence of host-symbiont interactions and their intertwining with host development, even in closely-related species.

Abbreviations: DAPI: 4',6-diamidino-2-phenylindole; FISH: Fluorescence *in situ* hybridization; T3SS: Type III secretion system.

ARTICLE HISTORY

Received 25 September 2020
Revised 19 October 2020
Accepted 19 October 2020

KEYWORDS

Symbiosis; bacteriome; metamorphosis; weevil; dryophthoridae; evolution; insect; rhynchophorus; nardonella; sitophilus

Main text

Symbiotic associations between animals and bacteria are present across all branches of the tree of life, in all ecological niches, and represent a major evolutionary driving force [1–3]. Symbiosis is particularly prevalent in insects, which often associate with intracellular bacteria (endosymbionts) that complement their diet and allow them to thrive on unbalanced niches, such as plant sap or cereals [4,5]. How endosymbiosis and host development are connected is a question of great interest to assess the full impact of bacterial partners on host biology [6]. In insects, a particularly important developmental period is the pupal stage: following larval development, holometabolous insects go through a complete internal and external morphological reorganization, metamorphosis, before emerging as adults [7,8]. Hence, endosymbiosis needs to be both maintained through this drastic morphological changes, and adapted to the physiological needs of the adult [9].

We recently addressed this question using the mutualistic association between the rice weevil *Sitophilus oryzae* and the bacterium *Sodalis*

pierantonius [10–12]. *S. pierantonius* is housed in specialized host cells, bacteriocytes, that group together in an organ, the bacteriome [4,12,13], and provides cereal weevils with vitamins and amino acids that are scarcely present in cereals [11,14,15]. Through fluorescent imaging techniques, we showed that, as the insect's internal organization changes during metamorphosis, the bacteriome is also completely remodeled [16]. The larval bacteriome, a unique organ located around the foregut, dissociates at the onset of metamorphosis, and bacteriocytes migrate along the midgut epithelium [16]. In pupae, bacteriocytes stop their migration around clusters of putative stem cells, positioned at regular intervals along the midgut [16]. At this stage, bacteria were seen entering uninfected stem cells, and electron microscopy further revealed the presence of *S. pierantonius* within the nuclei of these stem cells [16]. This was followed by the formation of multiple new bacteriomes at the apex of mesenteric ceca [16]. Using a dual-RNAseq strategy specifically tailored for the *Sitophilus-Sodalis* association, we showed host transcriptomic modifications in genes associated with cell adhesion

and motility, as well as cytoskeleton reorganization [16]. Furthermore, bacterial genes encoding the flagellum apparatus and the type III secretion system (T3SS) were upregulated in pupae [16]. This indicates that *S. pierantonius* transiently adopts an infectious behavior, potentially allowing the bacterium to infect stem cells and, transiently, their nuclei, and to trigger their differentiation into bacteriocytes. Interestingly, it was previously shown that, following metamorphosis, endosymbiont density massively increases during one week, whereupon endosymbionts and bacteriocytes are rapidly eliminated and recycled [14]. Endosymbiont growth was shown to match increasing needs for aromatic amino acids used by *S. oryzae* to rapidly build a strong cuticle following final ecdysis [14]. Hence, we hypothesized that the bacteriome reorganization during metamorphosis is a morphological optimization for the highly demanding and dynamic adult stage, allowing for the growth of the symbiotic population and more efficient host-symbiont metabolic exchanges. This morphological remodeling, synchronously driven by host and endosymbiont transcriptomic changes, shows the extent to which endosymbiosis is integrated with both host developmental processes and metabolic needs.

Whether bacteriome reorganization is conserved across species remains elusive. Data from two other *Sitophilus* species, *S. zeamais* and *S. granarius*, show that the process described in *S. oryzae* is conserved within these cereal-feeding species [14]. However, most histological data published so far on related insects are limited to larvae [17,18], and did not address the fate of symbiotic organs during metamorphosis. Here, we provide additional imaging data from a closely-related coleopteran of the Dryophthoridae family, the red palm weevil *Rhynchophorus ferrugineus*. The red palm weevil is one of the most damaging palm-trunk boring pest worldwide, and feeds mainly on sap and tender tissues [19]. *R. ferrugineus* harbors the endosymbiont *Nardonella*, the ancestral endosymbiont of the Dryophthoridae family to which *Sitophilus* weevils belong [17,20,21]. Wild *R. ferrugineus* were collected and fluorescence *in situ* hybridization (FISH) with a *Nardonella*-specific probe was conducted on dissected bacteriomes and guts. Surprisingly, while the gut underwent morphological reorganization during *R. ferrugineus*' metamorphosis, the bacteriome remained unchanged and unmoved, collar-shaped at the foregut-midgut junction, from larva to imago (Figure 1). This markedly contrasts with our previous observations in *Sitophilus*. Nonetheless, endosymbionts

were observed within the oocytes (Figure 2), similarly to *Sitophilus* [22], indicating that transmission and, potentially, bacteriome formation during embryogenesis, are conserved.

Both *Nardonella* and *S. pierantonius* are known to participate in host cuticle formation by providing aromatic amino acids [14,21], a feature that is shared across other beetle-bacteria associations [23,24]. In coleopterans, a thick and strong cuticle is essential to protect adults against predators, desiccation, and other environmental conditions [23]. However, the 'urgency' in cuticle formation that was shown in *S. oryzae* (*i.e.* a rapid increase in endosymbiont density associated with a rapid cuticle synthesis) has not been proven in *R. ferrugineus*. Previous work on the related weevil *Pachyrhynchus infernalis*, also harboring *Nardonella*, showed signs of cuticle synthesis 35 days after final ecdysis, as evidenced by changes in cuticle color and thickness [21], a much longer timespan than that observed in *S. oryzae* (*i.e.* 7 days, 14). A similar timespan (28 days) was observed for cuticle synthesis in *Pachyrhynchus sarcitis kotoensis* [25]. Interestingly, the cereal weevil *S. granarius* also exhibits a slower cuticle synthesis and symbiont dynamics than *S. oryzae* (~15 days until cuticle completion, versus 7 days in *S. oryzae*) [14]. This was correlated with a less contrasted symbiont dynamics in *S. granarius* associated with a 'flattened' endosymbiont density curve [14]. Unlike *S. oryzae* and *S. zeamais*, *S. granarius* is not found in field cereal crops but instead in grain store-rooms of temperate countries, and only possesses vestigial wings preventing this insect from flying between different sites [26]. It was hypothesized that this less demanding lifestyle in a relative stable environment does not require a rapid cuticle synthesis [14]. As *Pachyrhynchus* spp. are flightless [27] and *R. ferrugineus* shows limited dispersion in young adults [28], it is likely that these more sedentary lifestyles have resulted in relaxed constraints on the speed of cuticle synthesis following adult metamorphosis, similarly to *S. granarius*. Thus, we speculate that the low endosymbiont dynamics and density, and the reduced host-symbiont metabolic exchanges, might be adapted to the slower cuticle synthesis, and hence that a morphological reorganization of the bacteriome may not be needed in these species.

It is noteworthy that *Nardonella*'s reduced genome lacks the genes encoding the T3SS and the flagellum apparatus [21], and potentially the ability to infect new cells, which might be crucial for the

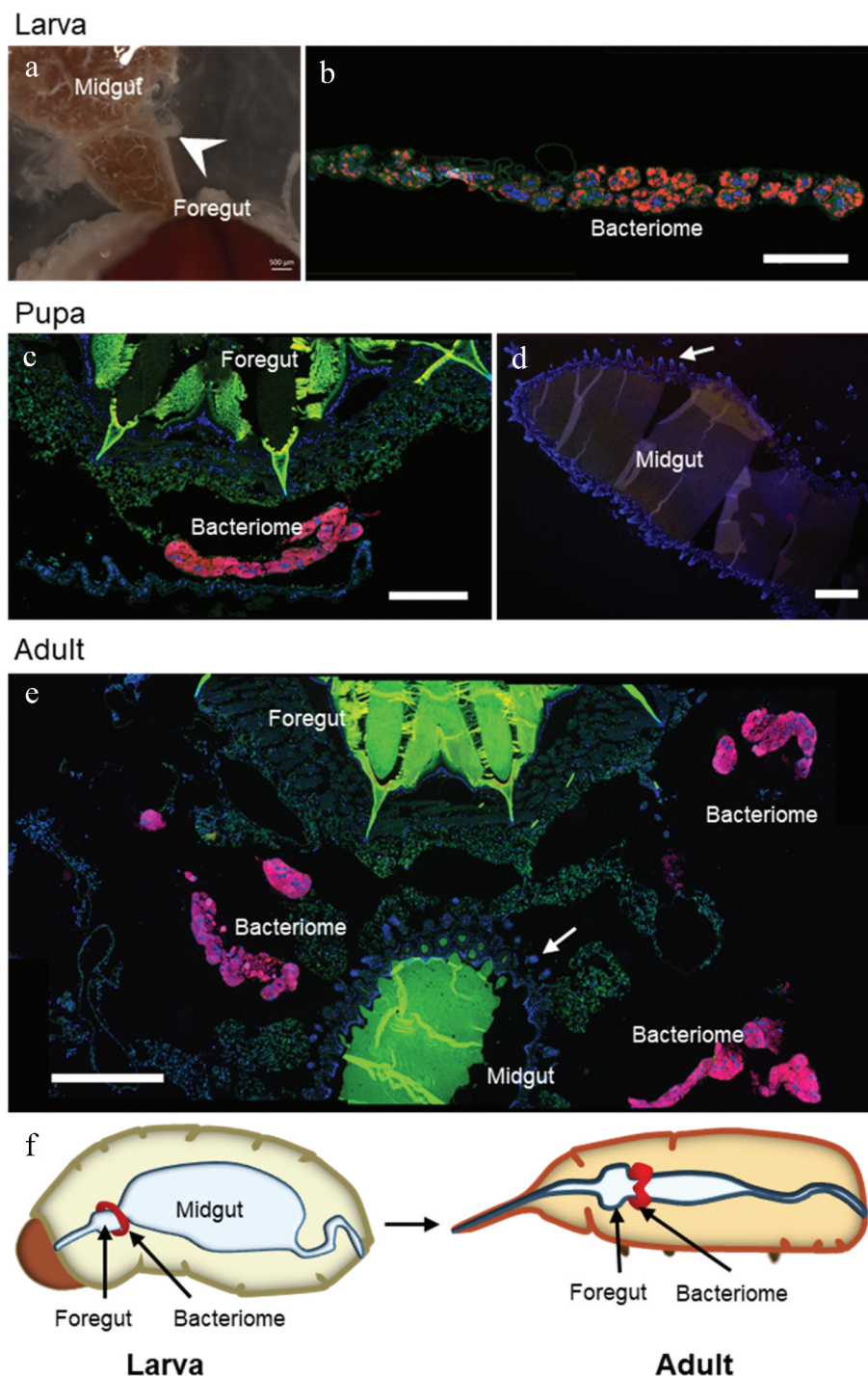


Figure 1. Bacteriome morphology throughout metamorphosis in *Rhynchophorus ferrugineus*, in the final larval instar (a-b), pupae (c-d), and young adults (e). **A:** Image of a dissected larval gut, with the collar-shaped bacteriome (white arrowhead) attached to the foregut-midgut junction. It can be seen detached and uncoiled in (b). **B-E:** Photos from FISH experiments targeting *R. ferrugineus*' endosymbiont, *Nardonella*. The bacteriome does not change in morphology or localization along metamorphosis: it remains a collar-shaped organ at the foregut-midgut junction. Mesenteric ceca can be seen forming in the midgut (white arrows in D, E), but no bacteriocyte is observed in these areas, unlike in *S. oryzae*. Red: *Nardonella*; green: autofluorescence; blue: DAPI. Scale bar: 250 μm for B, C; 500 μm for A, D, E. **F:** Schematic representation of the bacteriome in a larva and an adult.

morphogenesis of multiple bacteriomes in *Sitophilus* [16]. Nonetheless, we showed here that *Nardonella* is able to enter oocytes, suggesting the existence of other mechanisms enabling this endosymbiont to enter host

cells. Studies in other associations have indicated that various mechanisms have been selected along evolution ensuring endosymbiont transmission to the next generation. This includes exo-/endocytotic processes,

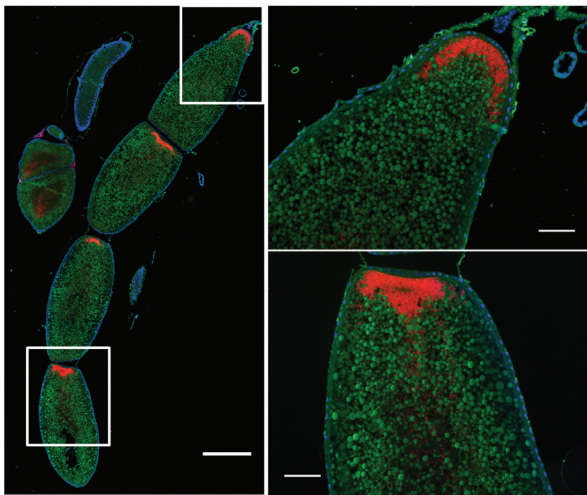


Figure 2. *Nardonella*-infected oocytes in *Rhynchophorus ferrugineus*. Red: *Nardonella*; Green: Autofluorescence; Blue: DAPI. The left image represents four oocytes in a female ovariole, observed in fluorescence microscopy following FISH treatment against the endosymbiont *Nardonella*. Scale bar is 0.5 mm in the left photo and 100 μ m in the close-ups.

which was described in the pea aphid/*Buchnera* association where *Buchnera* also lacks a T3SS [29], maternal bacteriocyte content transfer as observed in the ant *Cardiocondyla obscurior* [30], or the integration of a full, unique bacteriocyte inside the oocytes, as described in whiteflies [31,32]. From our observations, we can likely exclude the latter mechanism in the *R. ferrugineus*/*Nardonella* association; however further investigation will be required to test the potential implication of the two former mechanisms, or to highlight the selection of another mechanism in this association.

In conclusion, this work highlights the diversity of host-symbiont interactions and morphological features of endosymbiosis in insects of the same family, echoing previous work performed in closely-related stinkbugs [33]. Despite being phylogenetically very close, endosymbiosis in *S. oryzae* and *R. ferrugineus* shows drastically different developmental profiles: both bacteriome morphology and endosymbiont density are highly dynamic in *S. oryzae*, while *R. ferrugineus* exhibits higher stability. While the roots of these dissimilarities are unknown, a lower necessity for *R. ferrugineus* to quickly complete a cuticle is a hypothesis worth investigating. Detailed comparative work on symbiotic and asymptomatic coleopterans should be conducted to fully understand the extent of symbiotic contribution to cuticle synthesis and how it has impacted coleopteran evolution and diversification.

Material and methods

R. ferrugineus weevils were collected in the wild, on Italian palm trees. Both larvae and adults were kept on an apple diet at 28°C until dissection. Insect organs (bacteriomes, guts, ovaries) were dissected in sterile phosphate buffer saline and fixed in 4% paraformaldehyde. Samples were processed and sectioned for histological work as previously described [34].

FISH was performed as previously described [35]. Three individuals were observed per stage (larva, pupa, adult). Probe sequence for *Nardonella* was: TAMRA-CGCGAGAATGAGCAAATCTT (10 μ g/mL). Observations, image acquisition and treatment were performed as previously described [34]. Figure 1b-1e and left panel of Figure 2 are mosaics of several photos fused together using the ‘Stitching’ plugin in ImageJ.

Acknowledgments

Funding for this project was provided by Institut National de Recherche pour l’Agriculture, l’Alimentation et l’Environnement (INRAE), Institut National des Sciences Appliquées- Lyon (INSA-Lyon) BC thanks Suma Pompeo for his help with sampling.

Author contributions

JM, AZR, and AH designed research; JM collected tissues and conducted FISH experiments; BC provided living insects and assisted with tissue collection; JM, AZR, and AH wrote the manuscript.

Disclosure statement

The authors report no conflict of interest.

Funding

This work was supported by the Agence Nationale de la Recherche [ANR-17_CE20_0031_01, GREEN]; Institut National de Recherche pour l’Agriculture, l’Alimentation et l’Environnement (INRAE); Institut National des Sciences Appliquées- Lyon (INSA-Lyon).

References

- [1] McFall-Ngai M, Hadfield MG, Bosch TCG, et al. Animals in a bacterial world, a new imperative for the life sciences. *Proc Natl Acad Sci U S A.* 2013;110:3229–3236.
- [2] Moran NA. Symbiosis as an adaptive process and source of phenotypic complexity. *Proc Natl Acad Sci U S A.* 2007;8627–8633. DOI:10.1073/pnas.0611659104

- [3] Gilbert SF, Sapp J, Tauber AI. A symbiotic view of life: we have never been individuals. *Q Rev Biol*. 2012;87:325–341.
- [4] Buchner P. Endosymbiosis of animals with plant microorganisms. New York: Interscience Publishers/John Wiley; 1965.
- [5] Baumann P. Biology of bacteriocyte-associated endosymbionts of plant sap-sucking insects. *Annu Rev Microbiol*. 2005;59:155–189.
- [6] Gilbert SF, Bosch TCG, Ledón-Rettig C. Eco-Evo-Devo: developmental symbiosis and developmental plasticity as evolutionary agents. *Nat Rev Genet*. 2015;16:611–622.
- [7] Truman JW, Riddiford LM. The origins of insect metamorphosis. *Nature*. 1999;401:447–452.
- [8] Rolff J, Johnston PR, Reynolds S. Complete metamorphosis of insects. *Philos Trans R Soc Lond B Biol Sci*. 2019;374:20190063.
- [9] Hammer TJ, Moran NA. Links between metamorphosis and symbiosis in holometabolous insects. *Philos Trans R Soc B Biol Sci*. 2019;374:20190068.
- [10] Heddi A, Grenier AM, Khatchadourian C, et al. Four intracellular genomes direct weevil biology: nuclear, mitochondrial, principal endosymbiont, and Wolbachia. *Proc Natl Acad Sci U S A*. 1999;96:6814–6819.
- [11] Oakeson KF, Gil R, Clayton AL, et al. Genome degeneration and adaptation in a nascent stage of symbiosis. *Genome Biol Evol*. 2014;6:76–93.
- [12] Heddi A, Nardon P. *Sitophilus oryzae* L: A model for intracellular symbiosis in the Dryophthoridae weevils (Coleoptera). *Symbiosis*. 2005;39:1–11.
- [13] Mansour K. Preliminary studies on the bacterial cell-mass (accessory cell-mass) of *Calandra oryzae* (Linn.): the rice weevil. *Q J Microsc Sci*. 1930;2-73:421–435.
- [14] Vigneron A, Masson F, Vallier A, et al. Insects recycle endosymbionts when the benefit is over. *Curr Biol*. 2014;24:2267–2273.
- [15] Wicker C. Differential vitamin and choline requirements of symbiotic and aposymbiotic *S. oryzae* (Coleoptera: curculionidae). *Comp Biochem Physiol Part A Physiol*. 1983;76:177–182.
- [16] Maire J, Parisot N, Galvao Ferrarini M, et al. Spatial and morphological reorganization of endosymbiosis during metamorphosis accommodates adult metabolic requirements in a weevil. *Proc Natl Acad Sci*. 2020;117:19347–19358.
- [17] Conord C, Despres L, Vallier A, et al. Long-term evolutionary stability of bacterial endosymbiosis in curculionidae: additional evidence of symbiont replacement in the dryophthoridae family. *Mol Biol Evol*. 2008;25:859–868.
- [18] Nardon P, Lefèvre C, Delobel B, et al. Occurrence of endosymbiosis in dryophthoridae weevils: cytological insights into bacterial symbiotic structures. *Symbiosis*. 2002;33:227–241.
- [19] Faleiro JR. A review of the issues and management of the red palm weevil *Rhynchophorus ferrugineus* (Coleoptera: rhynchophoridae) in coconut and date palm during the last one hundred years. *Int J Trop Insect Sci*. 2006;26:135–154.
- [20] Lefèvre C, Charles H, Vallier A, et al. Endosymbiont phylogenesis in the dryophthoridae weevils: evidence for bacterial replacement. *Mol Biol Evol*. 2004;21:965–973.
- [21] Anbutsu H, Moriyama M, Nikoh N, et al. Small genome symbiont underlies cuticle hardness in beetles. *Proc Natl Acad Sci*. 2017;114:E8382–E8391.
- [22] Nardon P. Ovogenèse et transmission des bactéries symbiotiques chez le charançon *Sitophilus oryzae* L. (Coleoptera: curculionidae). *Ann la Société Entomol Fr*. 2006;42:129–164.
- [23] Engl T, Eberl N, Gorse C, et al. Ancient symbiosis confers desiccation resistance to stored grain pest beetles. *Mol Ecol*. 2018;27:2095–2108.
- [24] Hirota B, Okude G, Anbutsu H, et al. A novel, extremely elongated, and endocellular bacterial symbiont supports cuticle formation of a grain pest beetle. *MBio*. 2017;8:e01482-17.
- [25] Wang LY, Rajabi H, Ghoroubi N, et al. Biomechanical strategies underlying the robust body armour of an aposomatic weevil. *Front Physiol*. 2018;9:1410.
- [26] Longstaff BC. Biology of the grain pest species of the genus *Sitophilus* (Coleoptera: curculionidae): a critical review. *Prot Ecol*. 1981;3:83–130.
- [27] Yeh HY, Tseng HY, Lin CP, et al. Rafting on floating fruit is effective for oceanic dispersal of flightless weevils. *J Exp Biol*. 2018;221:jeb190488.
- [28] Ávalos JA, Marti-Campoy A, Soto A. Study of the flying ability of *Rhynchophorus ferrugineus* (Coleoptera: dryophthoridae) adults using a computer-monitored flight mill. *Bull Entomol Res*. 2014;104:462–470.
- [29] Koga R, Meng X-Y, Tsuchida T, et al. Cellular mechanism for selective vertical transmission of an obligate insect symbiont at the bacteriocyte-embryo interface. *Proc Natl Acad Sci U S A*. 2012;109:E1230–7.
- [30] Klein A, Schrader L, Gil R, et al. A novel intracellular mutualistic bacterium in the invasive ant *Cardiocondyla obscurior*. *Isme J*. 2016;10:376–388.
- [31] Xu X, Li N, Bao X, et al. Patterns of host cell inheritance in the bacterial symbiosis of whiteflies. *Insect Sci*. 2020;27:938–946.
- [32] Luan J-B, Shan H-W, Isermann P, et al. Cellular and molecular remodelling of a host cell for vertical transmission of bacterial symbionts. *Proc Biol Sci*. 2016;283:218–230.
- [33] Matsuura Y, Kikuchi Y, Hosokawa T, et al. Evolution of symbiotic organs and endosymbionts in lygaeid stinkbugs. *Isme J*. 2012;6:397–409.
- [34] Maire J, Vincent-Monégat C, Balmant S, et al. Weevil pgrp-lb prevents endosymbiont TCT dissemination and chronic host systemic immune activation. *Proc Natl Acad Sci U S A*. 2019;116:5623–5632.
- [35] Maire J, Vincent-Monégat C, Masson F, et al. An IMD-like pathway mediates both endosymbiont control and host immunity in the cereal weevil *Sitophilus* spp. *Microbiome*. 2018;6:6.